



# Proteins: Structure & Function

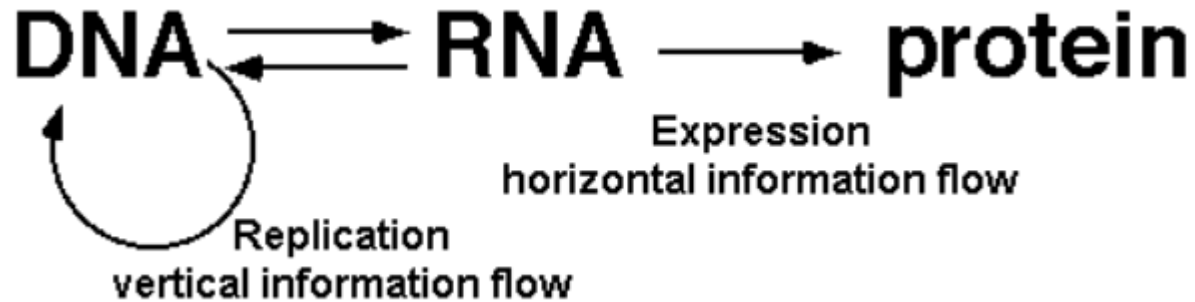
Ulf Leser

# This Lecture

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- Proteins
  - Structure
  - Function
  - Databases
- Predicting Protein Secondary Structure

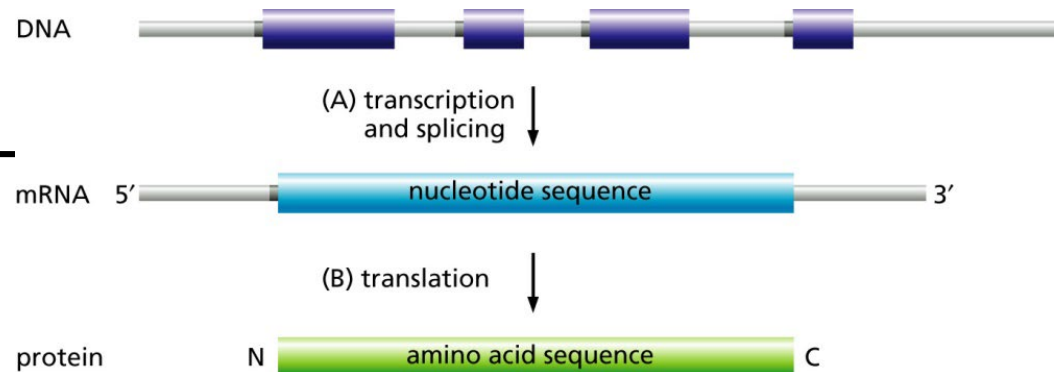
# Central Dogma of Molecular Biology



	U	C	A	G	
U	UUU Phenylalanine UUC	UCU Serine UCC UCA UCG	UAU Tyrosine UAC	UGU Cysteine UGC	U C A G
	UUA Leucine UUG		UAA Stop codon UAG Stop codon	UGA Stop codon UGG Tryptophan	
C	CUU Leucine CUC CUA CUG	CCU Proline CCC CCA CCG	CAU Histidine CAC	CGU Arginine CGC CGA CGG	U C A G
			CAA Glutamine CAG		
A	AUU Isoleucine AUC AUA	ACU Threonine ACC ACA ACG	AAU Asparagine AAC	AGU Serine AGC	U C A G
	AUG Methionine; initiation codon		AAA Lysine AAG	AGA Arginine AGG	
G	GUU Valine GUC GUA GUG	GCU Alanine GCC GCA GCG	GAU Aspartic acid GAC	GGU Glycine GGC GGA GGG	U C A G
			GAA Glutamic acid GAG		

# Details

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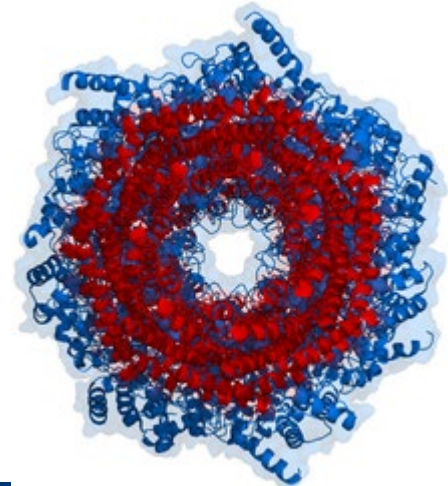
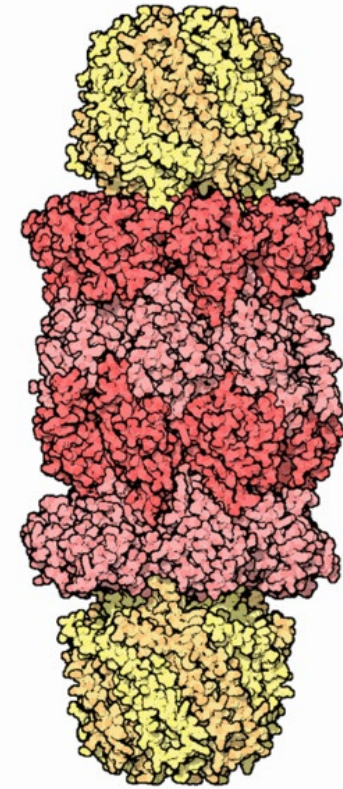


- Alternative Splicing
  - “One gene – one protein” is wrong
  - Exons may be spliced out from the mRNA
  - Human: at least **6 times more unique proteins** than genes
    - Also called isoforms
- Post-translational modifications
  - (De-)Phosphorylation, glycolysation, cleavage of signal peptides, ...
- Complexes: Proteins **physically and permanently grouping together** to perform a specific function

# Example Complex: Proteasome

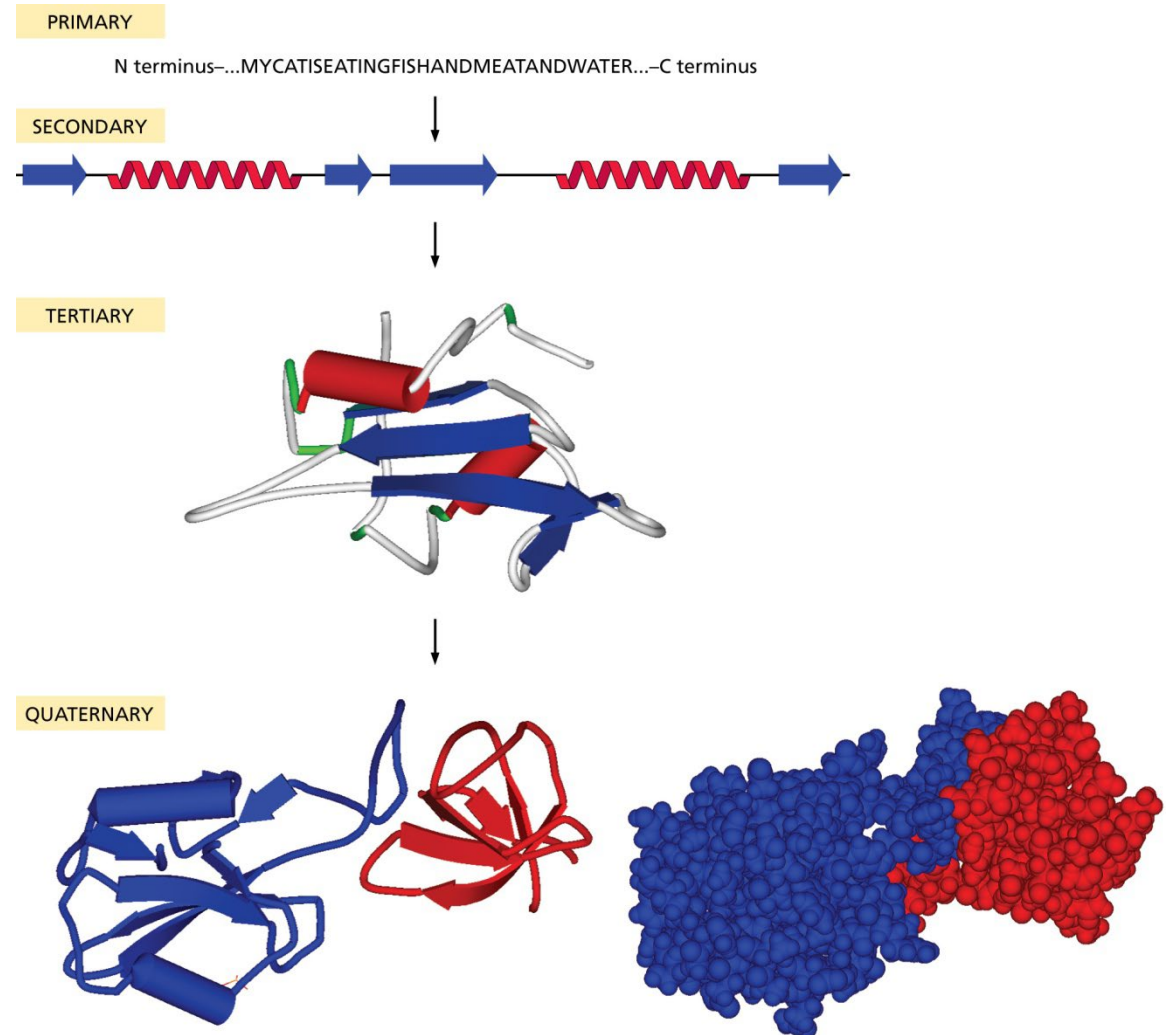
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- Function: Breaks (mis-folded, broken, superfluous, ...) proteins into small **peptides for reuse**
- Very large complex present in all eukaryotes (and more species)
  - >2000 kDa, consists of **dozens of individual proteins**
  - Formation of the complex is a complex process only partly understood yet



# Protein Structure

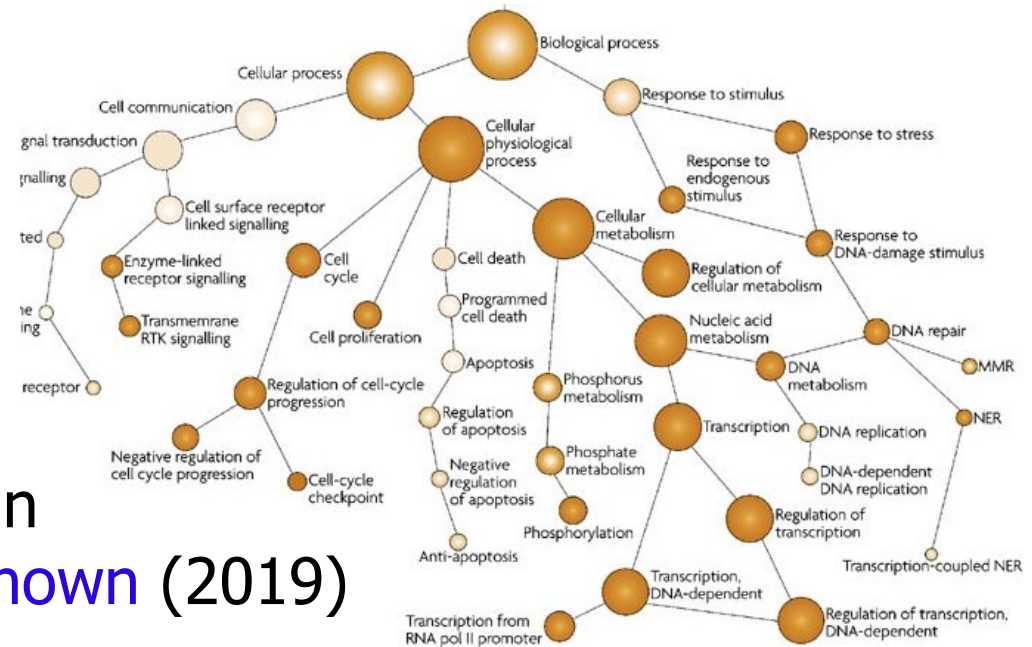
- Primary
  - 1D-Seq. of AA
- Secondary
  - 1D-Seq. of “subfolds”
- Tertiary
  - 3D-Structure
- Quaternary
  - Assembled complexes



# Protein Function

- Proteins perform many functions in living **organisms**

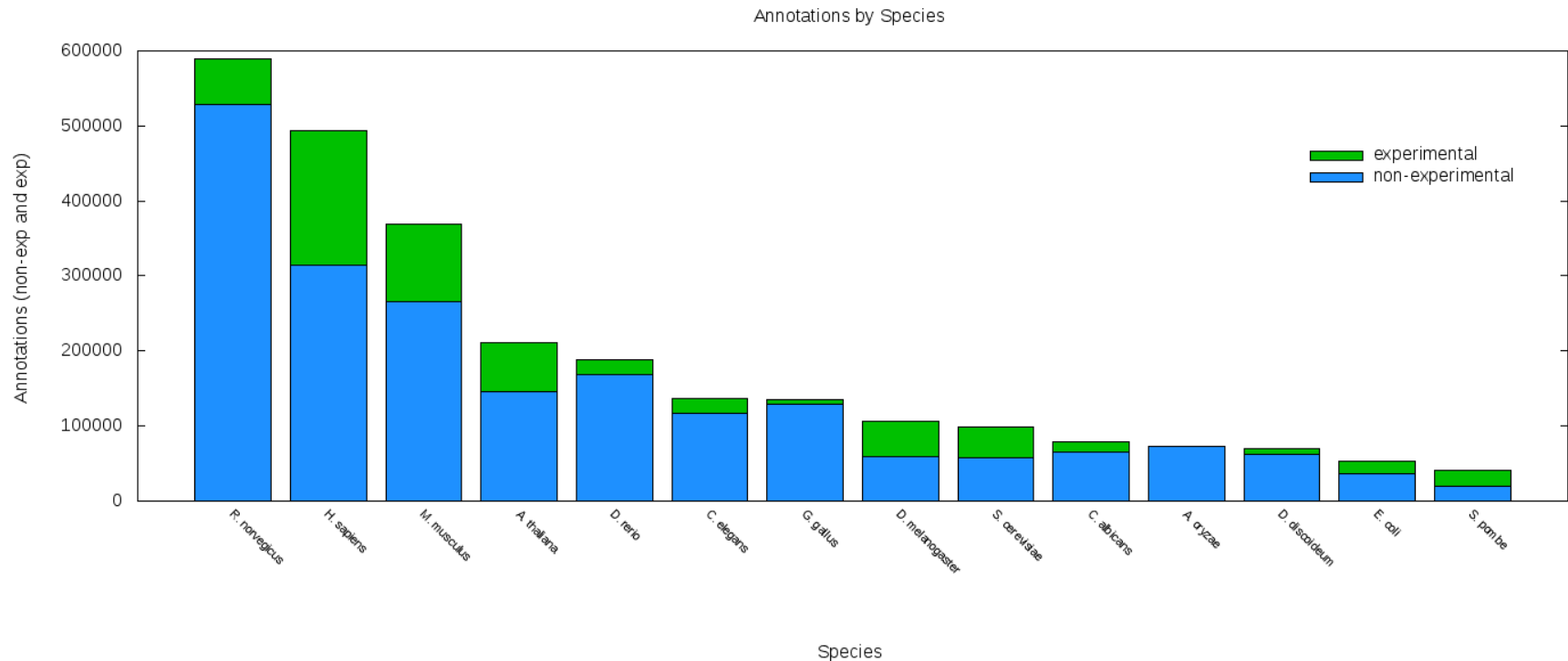
- Metabolism
- Signal processing
- Gene regulation
- Cell cycle
- ...



Nature Reviews | Cancer

- For  $\sim 20\%$  of all human gene, **no function is known** (2019)
- Describing function
  - **Gene Ontology**: 3 branches,  $>40.000$  concepts
  - Used world-wide to describe gene/protein function

# „Known“ Protein Functions

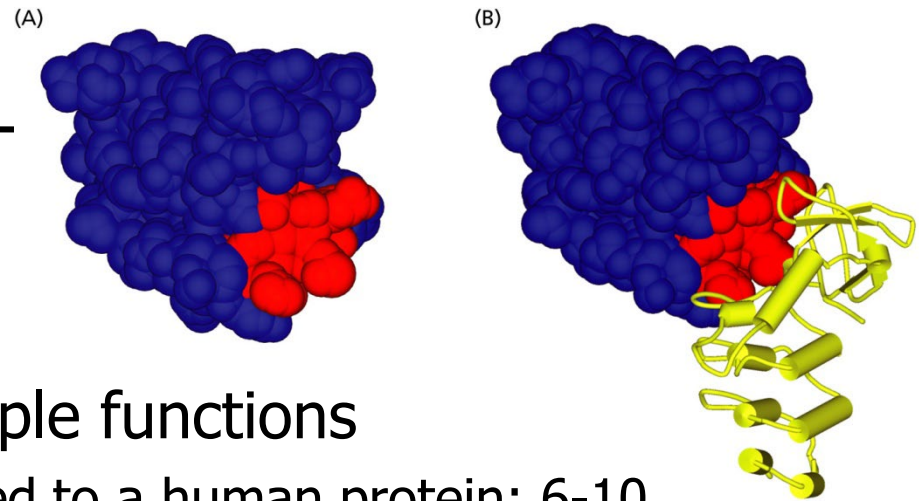


<http://geneontology.org/page/current-go-statistics>, June 2016



# Function and Motifs

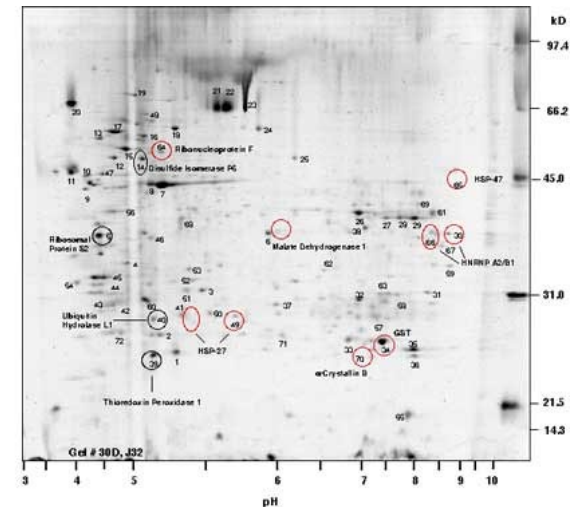
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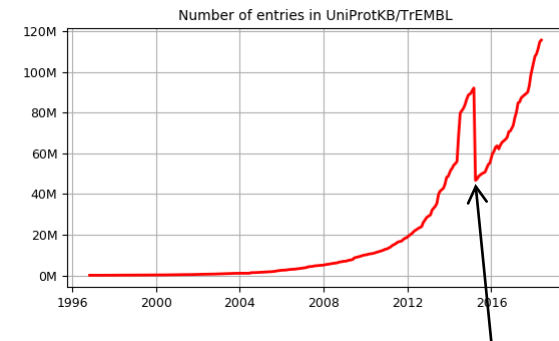
- Proteins usually have multiple functions
  - Avg. n# of GO terms assigned to a human protein: 6-10
- Functions are associated to **motifs or domains**
- There probably exist only 4000-5000 motifs
  - Proteins as assemblies of functional motifs
- Performing a function often requires **binding to another protein** or molecule
  - The binding requires a certain constellation of the protein structure
  - Major target of **pharmacological research**

# Proteomics – Large Scale Protein Identification

- Measuring gene expression: RNA-Seq, microarrays, PCR, ...
- Measuring **protein abundance** is much harder
  - **Isolating proteins** is very complex
  - Sequencing a protein is very slow
- Options (next lecture)
  - Isolation: 2D-Page, chromatography, ...
  - Identification: **Mass spectrometry**
  - De-novo sequencing with MS/MS
  - Quantification is very difficult

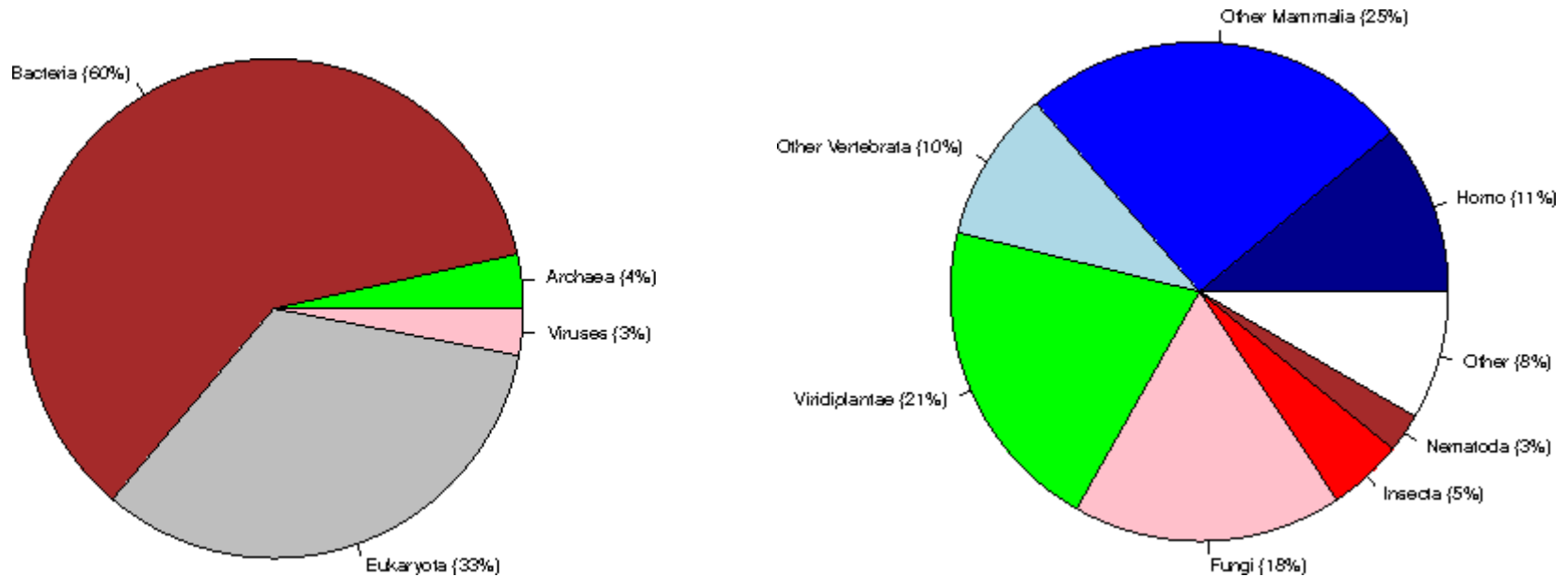


- “Standard” database for **protein sequences and annotation**
  - Original name: SwissProt
  - Started at the Swiss Institute of Bioinformatics, now mostly EBI
  - Other: PIR, HPRD
- Continuous growth and **curation**
  - >30 „Scientific Database Curators”
  - Quarterly releases
  - **Very rich set of annotations**
- Actually two databases
  - **SwissProt**: Curated, high quality, versioned
  - TrEMBL: Automatic generation from (putative) coding genomic sequences, low quality, redundant, much larger



Def. and removal of „redundant” sequences

# UniProt: Species [<http://www.expasy.org/sprot/relnotes/relstat.html>, June 2016]

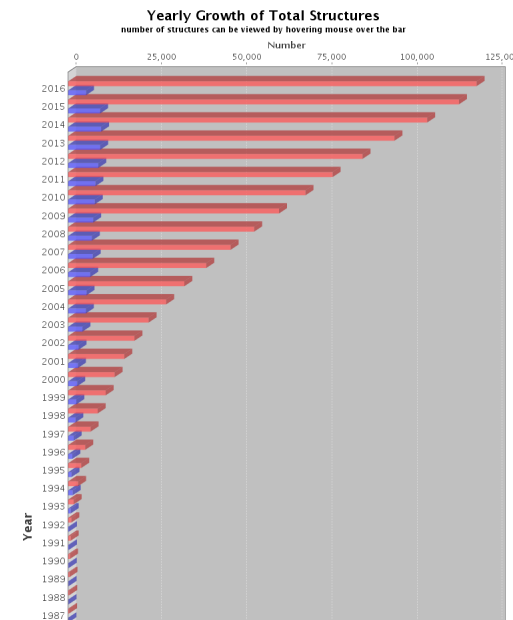


20258	Homo sapiens (Human)
16327	Mus musculus (Mouse)
9842	Arabidopsis thaliana (Mouse-ear cress)
7560	Rattus norvegicus (Rat)
6582	Saccharomyces cerevisiae (Baker's yeast)
5803	Bos taurus (Bovine)

...

# PDB – Protein Structure Database

- Oldest protein database, evolved from a book
- Experimentally determined **protein 3D-structures**
  - Plus some DNA, protein-ligand, complexes, ...
  - X-Ray (~75%), NMR (nuclear magnetic resonance, ~23%)
- Costly and **rather slow techniques**
  - Growth much smaller than that of sequence-related DBs
- Many problems with **legacy data** and data formats



<http://www.rcsb.org/pdb/statistics/contentGrowthChart.do?content=total>, June 2016

# This Lecture

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- Introduction
- Predicting Protein Secondary Structure
  - Secondary structure elements
  - Chou-Fasman
  - GOR IV
  - Other methods

# Amino Acids (AA)

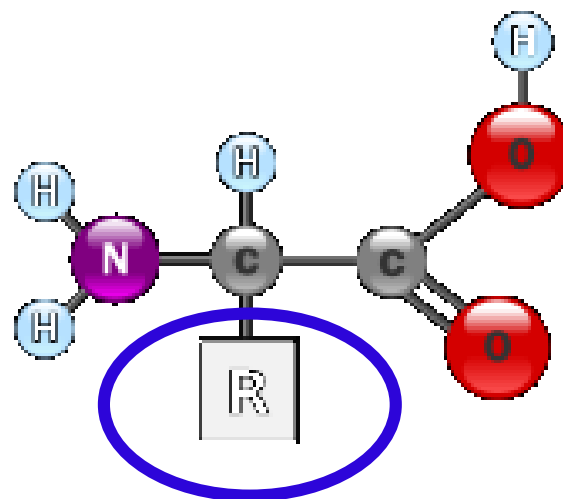
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- Amino acids: **Common core** and **specific residue**

- Core

- Amino group –  $\text{NH}_2$
- Central  $\text{C}_\alpha$  - Carbon – CH
- Carboxyl group –  $\text{COOH}$

- Residue: AA-specific



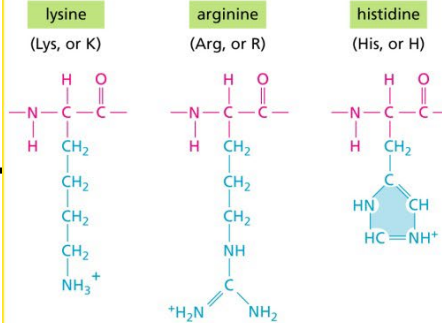
- Core: Chaining AA to protein sequences

- Residues (side chains): **Specific properties** of a AA

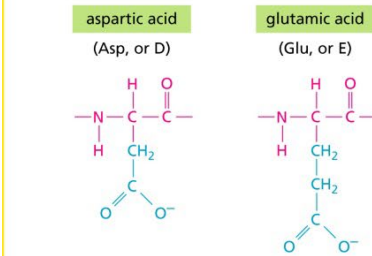
- Vary greatly between AA

# Side Chains

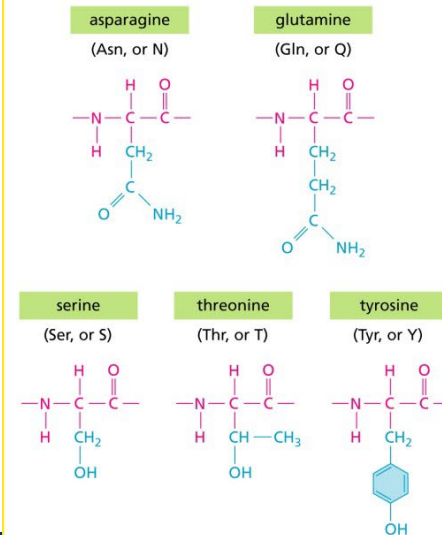
## BASIC SIDE CHAINS



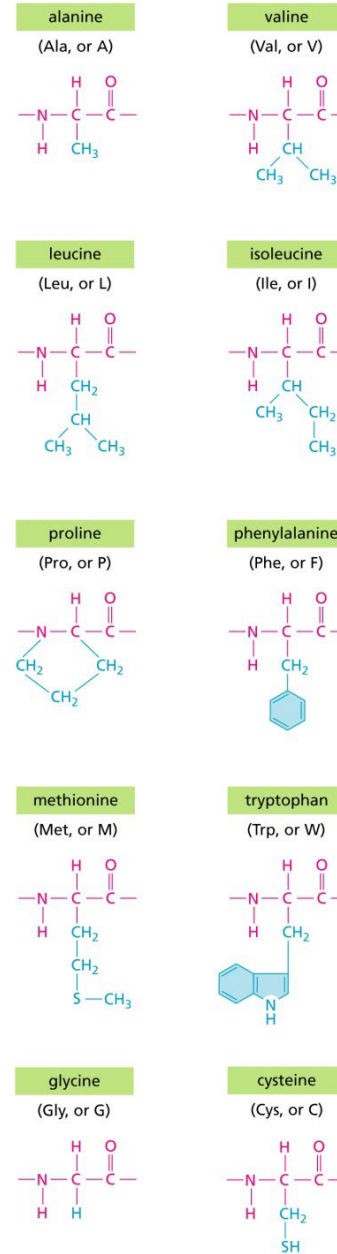
## ACIDIC SIDE CHAINS



## UNCHARGED POLAR SIDE CHAINS



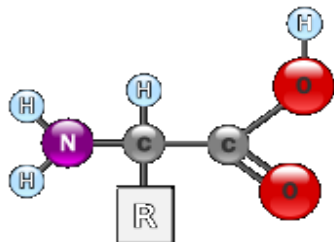
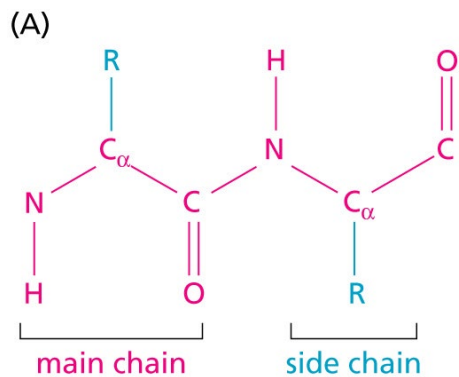
## NONPOLAR SIDE CHAINS



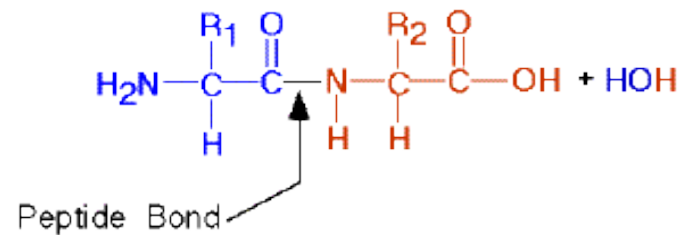
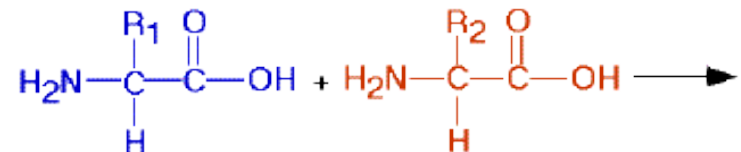


# Structure of a Protein

- Concatenation of **cores**: Backbone of AA chain (= protein)
  - Covalent **peptide bonds** between carboxyl and amino group
  - Loss of a H<sub>2</sub>O

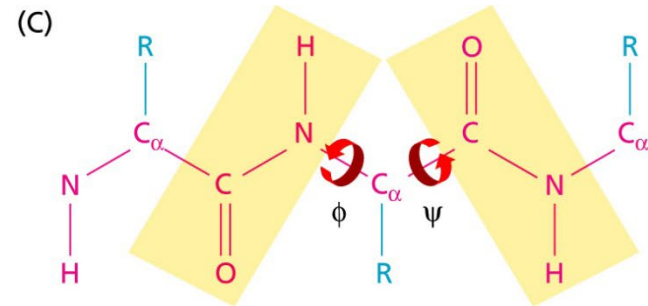
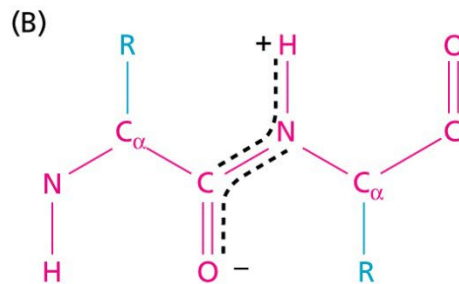
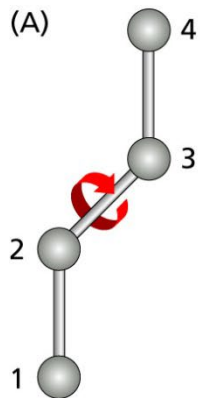


## Peptide Bond Formation



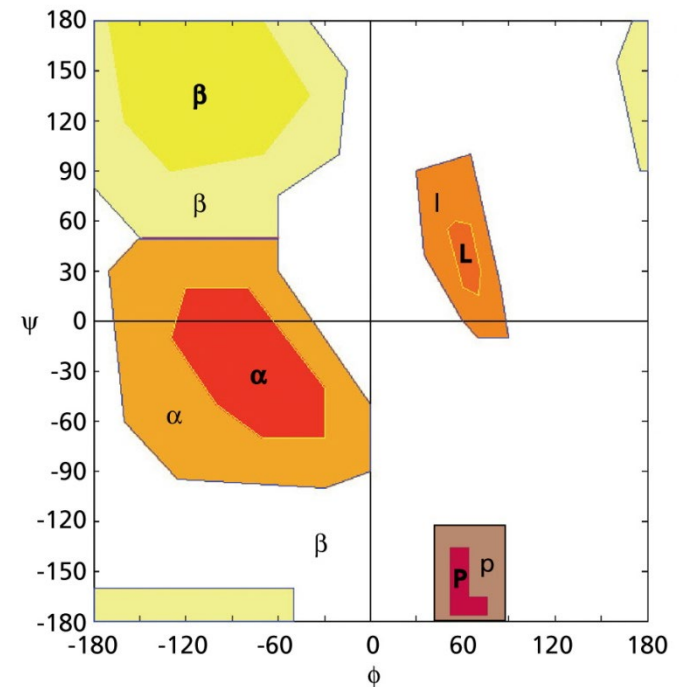
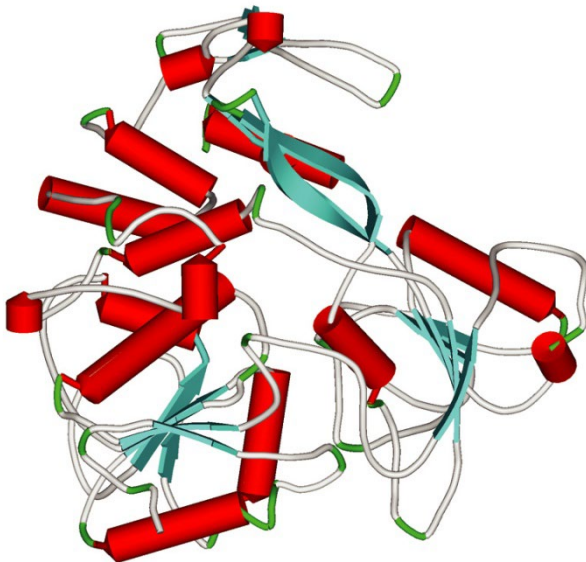
# Flexibility

- In principle, every chemical bond can rotate freely
  - Would allow arbitrary backbone structures
- In real proteins, observed angles are strongly constrained
  - Peptide bond (B) is “flat” – almost no torsion possible
  - Flexibility only in the  $C_\alpha$ -flanking bonds  $\phi$  and  $\psi$

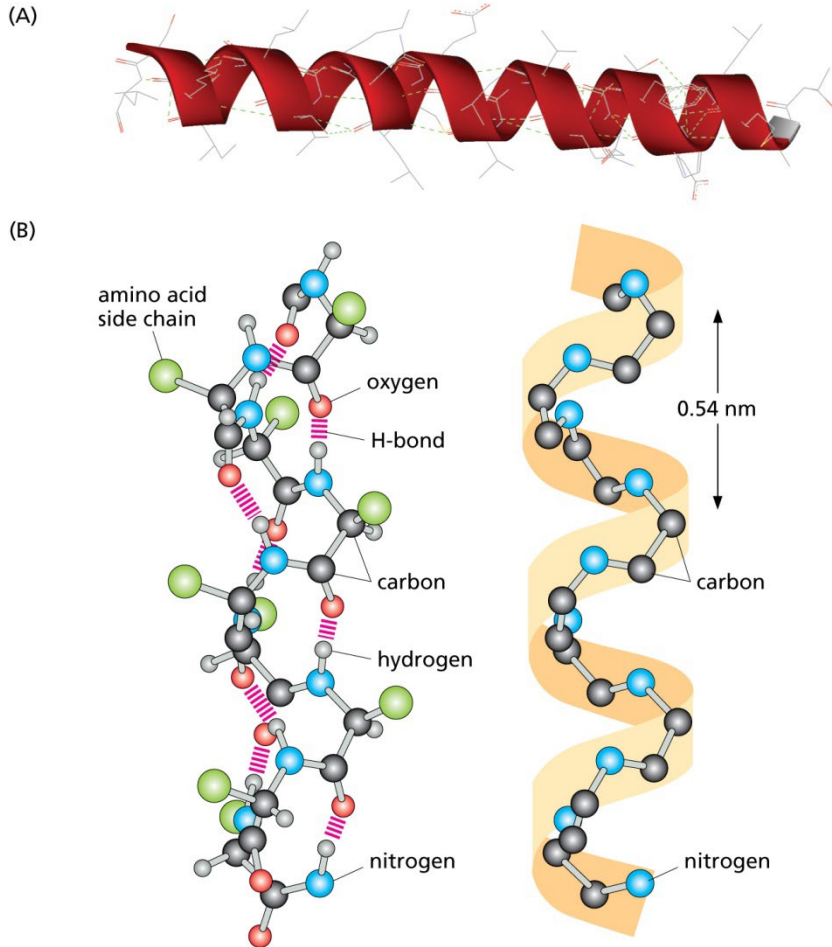


# Ramachandran Plots

- Combinations of  $\phi$  and  $\psi$  are **highly constrained**
  - Due to chemical properties of the backbone / side chains
- Two combinations are favored:  **$\alpha$ -helixes** and  **$\beta$ -sheets**
  - More detailed classifications exist
  - Angels lead to specific 3D structures
  - **Secondary structure**



# $\alpha$ -Helix

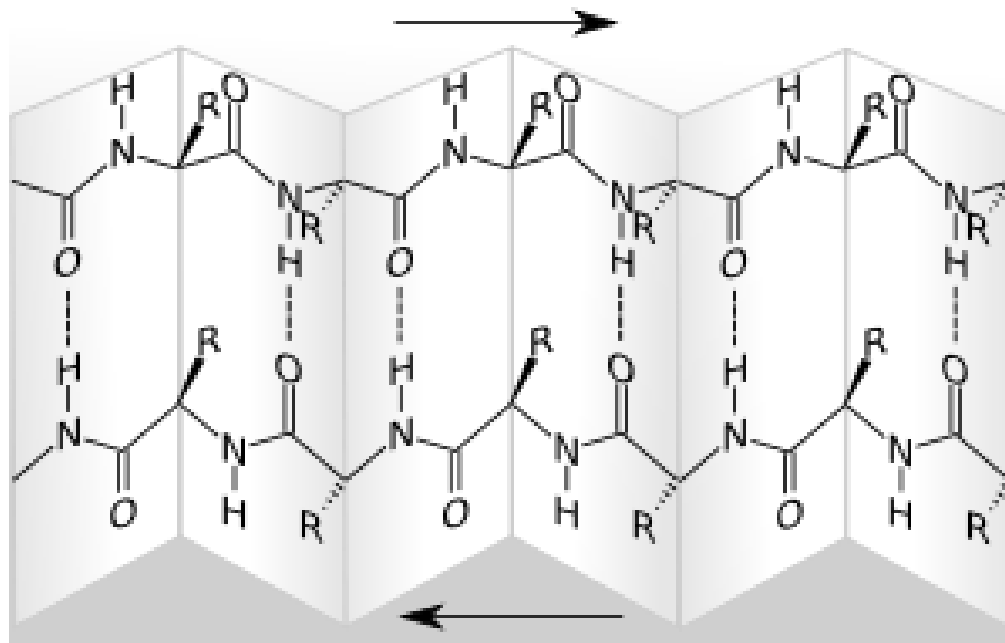


- Sequence of angles forming a regularly structured **helix**
- Additional bonds between amino and carboxyl groups
  - Very **stable structure**
- May have two orientations
  - Most are right-handed
- 3.4 AA per twist
- Often short, sometimes very long

# $\beta$ -Sheet

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- Two linear and **parallel stretches** ( $\beta$ -strands)
- Strands are bound together by hydrogen bonds
- Can be parallel or anti-parallel (wrt. N/C terminus)

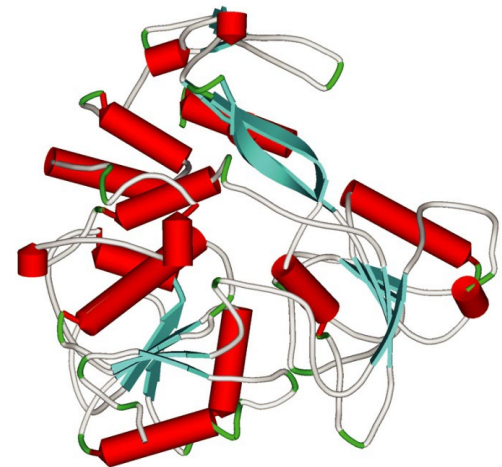


Quelle: Wikipedia

# Other Substructures

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- $\alpha$ -helices and  $\beta$ -sheets cover 50-80% of most proteins
- Other parts are called **loops** or **coils**
  - Usually less important for the structure of the protein
  - But **very important for its function**
  - Often exposed on the surface
  - Determine **binding** to other molecules



# Importance of Secondary Structure Prediction (SSP)

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- Secondary structure elements (SSE) are vital for the overall structure of a protein
- Often **evolutionary well conserved**
- SSE can be used to classify proteins
  - Mostly alpha, mostly beta, ...
  - Such classes are highly correlated with function
- SSE gives important **clues to protein structure**
- SSP **much simpler** than 3D structure prediction
  - And 3D structure prediction can benefit a lot from a good SSP

# Predicting Secondary Structure

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- SSP: Given a protein sequence, assign each AA in the sequence to one of the three classes Helix (H), Strand (E), or Coil (-)

```
KVYGRCELAAAMKRLGLDNYRGYSLGNWVCAAKFESNFNTHATNRNTD
GSTDYGILQINSRWWCNDGRTPGSKNLCNIPCSALLSSDITASVNCAK
KIASGGNGMNAWVAWRNRCKGTDVHAWIRGCRL
```



```
KVYGRCELAAAMKRLGLDNYRGYSLGNWVCAAKFESNFNTHATNRNTD
----HHHHHHHH-----EEEE-----HHHHHHHH--
GSTDYGILQINSRWWCNDGRTPGSKNLCNIPCSALLSSDITASVNCAK
----EEEEEEEEEEEEEEEE-----HHHHHH
KIASGGNGMNAWVAWRNRCKGTDVHAWIRGCRL
HHH-----EEE-----EEEE----
```



# Classification

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- **Classification**: Classify each AA into one of **three classes**
- Classification is a **fundamental problem**
  - Classify the readout of a microarray as diseased / healthy
  - Classify a subsequence of a genome as coding / non-coding
  - Classify an email as spam / no spam
- Many **different techniques**: Naïve Bayes, Regression, Decision Trees, SVMs, Neural Networks, ...
  - **Classification function** learned from properties of known objects
  - Often use same representation (feature vectors) of objects – methods exchangeable
- The following is a heuristic approach
  - Simple to explain, classical, no ML required, not too bad

# This Lecture

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- Introduction
- Predicting Protein Secondary Structure
  - Secondary structure elements
  - Chou-Fasman
  - Other methods

# Chou-Fasman Algorithm

Chou & Fasman (1974). Prediction of protein conformation. Biochemistry 13

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- Observation: Different AA favor different folds
  - Different AA are more or less often in H, E, C
  - Different AA are more or less often within, starting, or ending a stretch of H, E, C
- Chou-Fasman algorithm (rough idea)
  - Compute a score for the probability of any AA to be E / H
    - When both are improbable: Assign C
  - Basis: Relative frequencies in a set of sequences with known SSE
  - First assign each AA its most frequent class
  - Then perform several heuristic tricks to change classes
    - E.g. minimal length of stretches
    - Example: CCEEEEEECCCECE, not CCEEECEECCCECE

# Details [sketch, some heuristics omitted]

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- Let  $f_{j,k}$  be the relative frequency of observing AA  $j$  in class  $k$
- Let  $f_k$  be the average over all 20  $f_{j,k}$  values
- Compute the propensity  $P_{j,k}$  of AA  $j$  to be part of class  $k$  as

$$P_{j,k} = f_{j,k} / f_k$$

- This is not a **probability**, rather an odds-score
- Using  $P_{j,k}$ , classify each AA  $j$  for every class  $k$  into
  - Strong, normal, weak **builder** ( $H_{\alpha}, h_{\alpha}, I_{\alpha}, H_{\beta}, h_{\beta}, I_{\beta}$ )
    - Tendency to build a SS-element
  - Strong, weak **breaker** ( $B_{\alpha}, b_{\alpha}, B_{\beta}, b_{\beta}$ )
    - Tendency to stop a SS-element
  - Indifferent ( $i_{\alpha}, i_{\beta}$ )
  - Thus, we actually have 12 (13) classes

# Concrete Values

- Originally computed on only 15 proteins (1974)
- Read
  - Glu(tamate) often is at the start of a helix and often at the end of a strand
  - Met(hionine) often starts strands and regularly starts helices
  - ...

AS	$P_\alpha$	Klasse	AS	$P_\beta$	Klasse
Glu	1.53	H <sub>α</sub>	Met	1.67	H <sub>β</sub>
Ala	1.45		Val	1.65	
Leu	1.34		Ile	1.60	
His	1.24	h <sub>α</sub>	Cys	1.30	h <sub>β</sub>
Met	1.20		Tyr	1.29	
Gln	1.17		Phe	1.28	
Trp	1.14		Gln	1.23	
Val	1.14		Leu	1.22	
Phe	1.12		Thr	1.20	
Lys	1.07		I <sub>α</sub>	Trp	

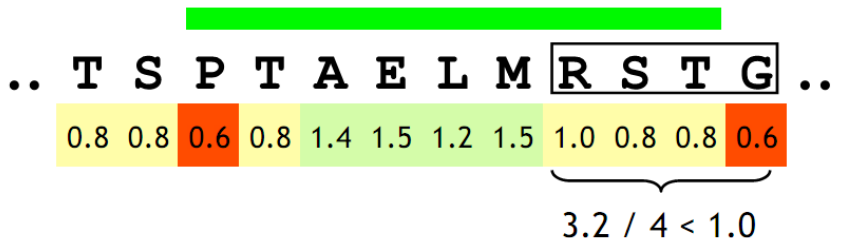
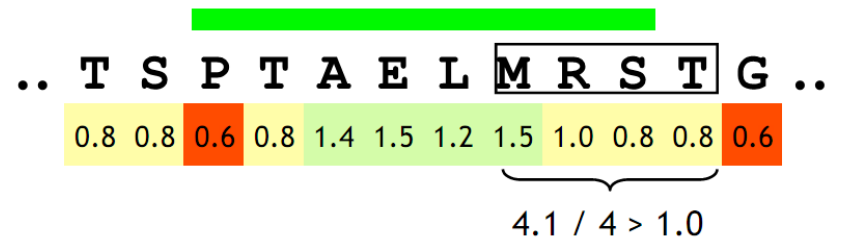
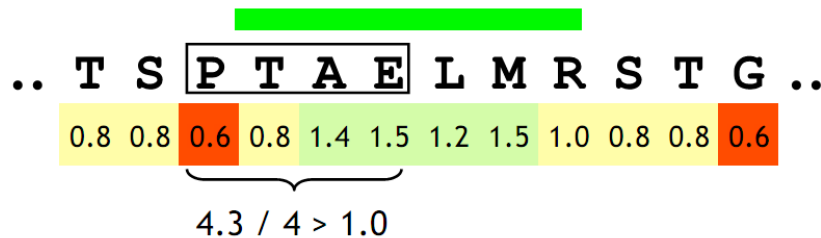
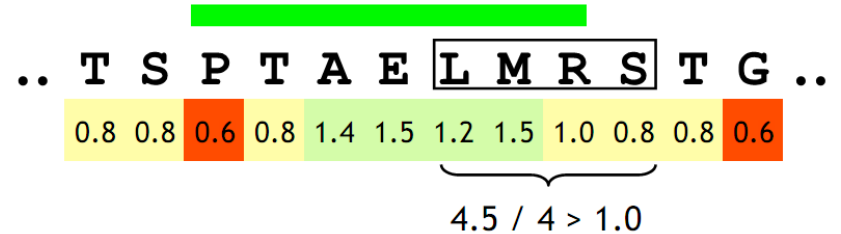
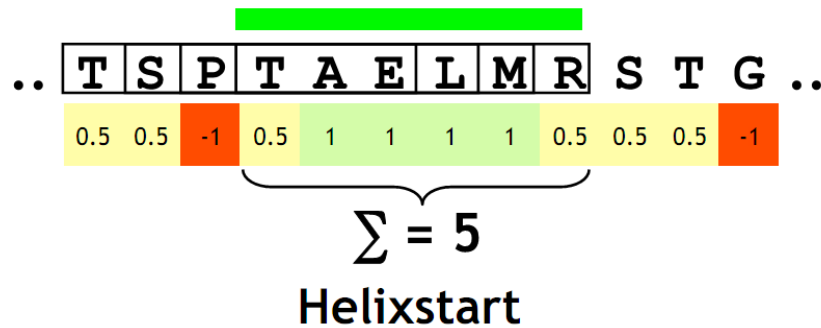
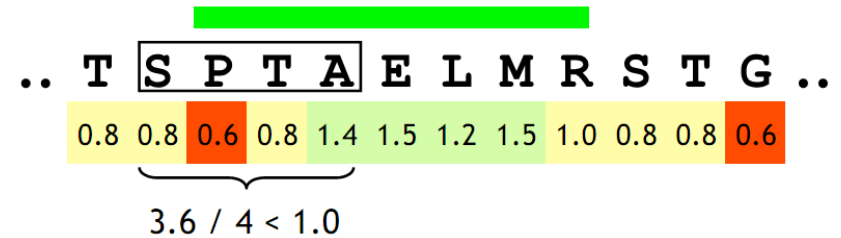
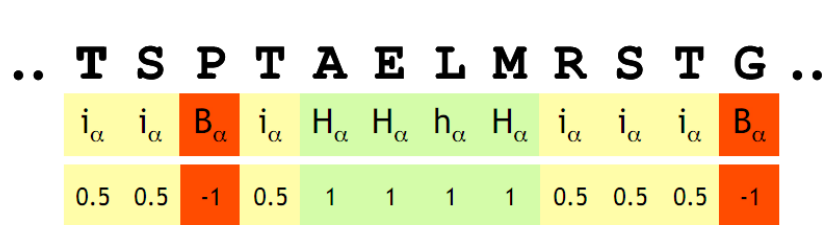
AS	$P_\alpha$	Klasse	AS	$P_\beta$	Klasse
Ile	1.00	I <sub>α</sub>	Ala	0.93	I <sub>β</sub>
Asp	0.98	i <sub>α</sub>	Arg	0.90	i <sub>β</sub>
Thr	0.82		Gly	0.81	
Ser	0.79		Asp	0.80	
Arg	0.79	b <sub>α</sub>	Lys	0.74	b <sub>β</sub>
Cys	0.77		Ser	0.72	
Asn	0.73		His	0.71	
Tyr	0.61		Asn	0.65	
Pro	0.59	B <sub>α</sub>	Pro	0.62	B <sub>β</sub>
Glu	0.53		Glu	0.26	

# Algorithm for Helices

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- Score each AA with 1 ( $H_\alpha, h_\alpha$ ), 0.5 ( $I_\alpha, i_\alpha$ ), or -1 ( $B_\alpha, b_\alpha$ )
  - Heuristic discretization – don't trust your counts too much
- Find **helix cores**: subsequences of length 6 with an aggregated AA score  $\geq 4$
- Starting from the middle of each core, shift a **window of length 4** to the left, then to the right
  - Compute aggregated score A using original  $P_{j,k}$  values inside each window
  - If  $A \geq 4$ , continue the helix; otherwise stop
- Similar method for strands
- **Conflicts** (regions assigned both H and E) are resolved based on higher aggregated score

# Example [Source: O. Kohlbacher, "Strukturvorhersage"]



# Performance

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- Accuracy app. 50-60%
  - Measured on per-AA correctness
- Prediction is **more accurate in helices** than in strands
- General problem of Chou-Fasman
  - Secondary structure is not only a local problem
  - Looking **only at single AAs** is not enough
    - Note: Scores are based on individual AA; aggregation by summation assumes **statistical independence** of pairs, triples ... in a class
- One needs to include the **context of an AA**



# This Lecture

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- Introduction
- Predicting Protein Secondary Structure
  - Secondary structure elements
  - Chou-Fasman
  - Other methods

# Classes of Methods

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- First generation: Properties of single AA only
  - Accuracy: 50-60%, e.g. Chou-Fasman (1974)
- Second generation: Include info. **about neighborhood**
  - Accuracy: ~65%, e.g. GOR (1974 – 1987)
- Third generation: Include info. from **homologous seq's**
  - Accuracy: ~70-75%, w.g. PHD (1994)
- Forth generation: Build **ensembles** of good methods
  - Accuracy: ~80%, e.g. Jpred (1998)
- Current performance
  - Jpred 4 (2015): 82% overall, ~90% for certain other properties
  - Spine-X (2012): 84% overall

# Further Reading

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- Gerhard Steger (2003). "Bioinformatik – Methoden zur Vorhersage von RNA- und Proteinstrukturen", Birkhäuser, chapter 8,10,11,13
- Many figures from Zvelebil, M. and Baum, J. O. (2008). "Understanding Bioinformatics", Garland Science, Taylor & Francis Group, chapter 2, 11, 12 (partly)
- Many examples from O. Kohlbacher, Vorlesung Strukturvorhersage, WS 2004/2005, Universität Tübingen